



## 🖳 Tag Pol Hot Start - Ab+ dual buffer

Heat-activatable DNA polymerase for high specificity, antibody-blocked Thermus aquaticus, recombinant, F coli

| Cat. Nº.    | Amount              |
|-------------|---------------------|
| D POL-124XS | 250 units           |
| POL-124S    | 500 units           |
| D POL-124M  | 1.000 units         |
| D POL-124L  | 2 x 1.000 units (M) |
| D POL-124XL | 4 x 1.000 units (M) |

## Unit Definition:

One unit is defined as the amount of the enzyme required to catalyze the incorporation of 10 nmoles of dNTPs into an acidinsoluble form in 30 minutes at 70 °C.

#### **Concentration:**

5 units/µL

Shipping:

Shipped on blue ice

## **Storage Conditions:**

Store at -20 °C

For in vitro use only!

### **Additional Storage Conditions:**

Avoid freeze/thaw cycles

## Shelf Life:

12 months

### **Kit contents:**

### Tag Pol Hot Start (blue cap)

5 units/µl Taq Pol Hot Start (#POL-123) in Tris-HCl pH 8.0 (25°C), KCl, EDTA, DTT, 50% (v/v) Glycerol, and stabilizers.

### Taq HS Reaction Buffer (red cap) - 5x conc.

Proprietary formulation supplied at pH 8,5 (25°C). Does not contain dyes or magnesium.

## Tag HS Reaction Buffer Green (red cap) - 5x conc.

Is supplemented with tracking dyes for direct loading of PCR products on gels. Does not contain magnesium.

## MgCl<sub>2</sub> Stock Solution (yellow cap)

50 mM MgCl<sub>2</sub>.

## **Description:**

Hot Start Pol provides improved specificity and sensitivity when amplifying low-copy-number targets in complex backgrounds or when prolonged room-temperature set up is required. The polymerase activity is blocked at ambient temperature and switched on automatically at the onset of the initial denaturation. The thermal activation prevents the extension of nonspecifically annealed primers and primer-dimer formation at low temperatures during PCR setup. The enzyme catalyzes the polymerization of nucleotides into duplex DNA in 5' $\rightarrow$ 3' direction in the presence of magnesium. It also possesses a  $5' \rightarrow 3'$ polymerization-dependent exonuclease replacement activity but lacks a  $3' \rightarrow 5'$  exonuclease activity.

### Activation step:

Hot Start Pol requires no prolonged heating or denaturing step. The polymerase inhibiting antibody is quickly released at the increased temperature of thermal cycling.

#### **PCR Reaction Setup**

The PCR procedure below shows appropriate volumes for a single 50-µL reaction. For multiple reactions, prepare a master mix of components common to all and then dispense appropriate volumes into each PCR reaction tube prior to adding template DNA and primers.

Thaw, mix, and briefly centrifuge each component before use.

Add the following components to a microcentrifuge tube:

#### 1. Prepare PCR master mix

Note: Consider the volumes for all components listed next steps to determine the correct amount of water required to reach your final reaction volume

| Components                 | 50 μL rxn     | [ final ]             |
|----------------------------|---------------|-----------------------|
| Water, grade PCR           | To 50 μL      |                       |
| 5x Reaction Buffer         | 10 µL         | 1X                    |
| dNTP (Mix 10 mM)           | 1 µL          | 200 µM                |
| Taq Pol Hot Start (5U/ μL) | 0,25 - 0,5 µl | 1,25 - 2,5 U/reaction |
| MgCl <sub>2</sub>          | 2 µl          | 2 mM                  |

Mix and briefly centrifuge the components.

### 2. Add template DNA and primers

| Components             | 50 µL rxn    | [final]        |
|------------------------|--------------|----------------|
| Foward primer (10 µM)  | 0,5 - 2,5 µl | 0,1 – 0,5 μM   |
| Reverse primer (10 µM) | 0,5 - 2,5 µl | 0,1 – 0,5 μM   |
| DNA template           |              | 10 pg – 1 µg** |

\*\*genomic DNA: 1 ng-1µg; plasmidial or viral DNA: 1 pg-1 ng

Cap each tube, mix, and briefly centrifuge the content.



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**DATA SHEET** 

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## 3. Optimization of MgCl<sub>2</sub> concentration:

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The 5x reaction buffer does not contain MgCl<sub>2</sub>. For an individual optimization, add MgCl<sub>2</sub> stock solution as shown in the table below:

| MgCl <sub>2</sub> Final Concentration   | 2,5 mM | 3 mM | 4 mM |
|---|--------|------|------|
| MgCl <sub>2</sub> stock volume to 50 µl | 2,5 µl | 3 µl | 4 µl |

## 4. Incubate reactions in a thermal cycler.

Recommended cycling conditions:

| Step                       |                         | Temp.    | Time        |
|----------------------------|-------------------------|----------|-------------|
| Initial denaturation       |                         | 95 °C    | 2 - 5 min   |
| 30 cycles                  | Denaturation            | 95 °C    | 15 - 30 sec |
|                            | Annealing <sup>1</sup>  | 45-68 °C | 15 - 30 sec |
|                            | Elongation <sup>2</sup> | 72 °C    | 1 min/kbp   |
| Final extension (optional) |                         | 72 °C    | 1 - 2 min   |
| Hold                       |                         | 4 - 8 °C |             |

1)The annealing temperature depends on the melting temperature of the primers used.

2)The elongation time depends on the length of the fragments to be amplified. A time of 1 min/kb is recommended.

For optimal specificity and amplification an individual optimization of the recommended parameters may be necessary for each new template DNA and/or primer pair.

